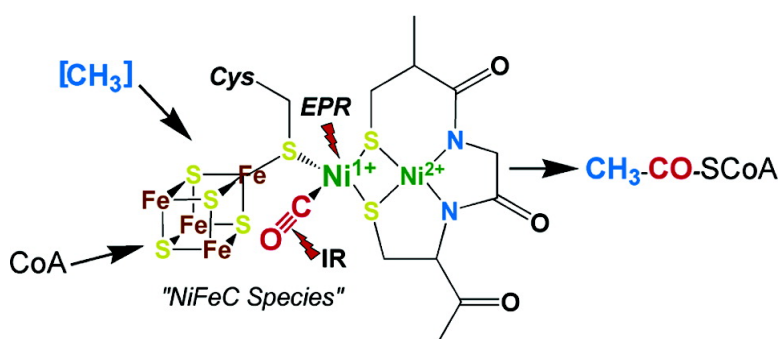


EPR and Infrared Spectroscopic Evidence That a Kinetically Competent Paramagnetic Intermediate is Formed When Acetyl-Coenzyme A Synthase Reacts with CO

Simon J. George, Javier Seravalli, and Stephen W. Ragsdale

J. Am. Chem. Soc., 2005, 127 (39), 13500-13501 • DOI: 10.1021/ja0528329 • Publication Date (Web): 07 September 2005

Downloaded from <http://pubs.acs.org> on March 25, 2009



More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Links to the 6 articles that cite this article, as of the time of this article download
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

[View the Full Text HTML](#)

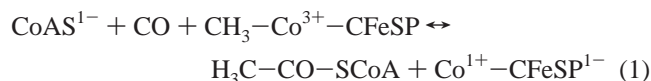
EPR and Infrared Spectroscopic Evidence That a Kinetically Competent Paramagnetic Intermediate is Formed When Acetyl-Coenzyme A Synthase Reacts with CO

Simon J. George,[†] Javier Seravalli,[‡] and Stephen W. Ragsdale^{*‡}

*Physical Biosciences Division, Lawrence Berkeley National Laboratory, Berkeley, California 94720, and
Department of Biochemistry, University of Nebraska, Lincoln, Nebraska 68588*

Received May 1, 2005; E-mail: sragdale1@unl.edu

CO dehydrogenase/acetyl-CoA synthase (CODH/ACS¹) is a bifunctional enzyme that enables archaea and bacteria to grow autotrophically on CO and H₂/CO₂ using the Wood–Ljungdahl pathway.^{2,3} The best characterized CODH/ACS from *Moorella thermoacetica*^{2,4} is an α₂β₂ heterotetrameric protein in which the β subunit (CODH) catalyzes the reduction of CO₂ to CO, while the α subunit (ACS) catalyzes the synthesis of acetyl-CoA (1) from Coenzyme A, CO, and a methyl cation donated by a methylated corrinoid iron–sulfur protein (CH₃–Co³⁺–CFeSP).⁵



The X-ray structures of all ACS proteins reveal an active site A-cluster that contains an [Fe₄S₄] cubane bridged by a cysteine sulfur atom to a binuclear center, which is composed of two Ni atoms.^{6,7} The proximal Ni (Ni_p), is attached to the cluster by a cysteinyl sulfur and to the distal Ni (Ni_d) by two additional cysteine residues. Ni_d, which assumes a square-planar geometry, is coordinated by the two bridging Cys and two backbone amide nitrogens.

In the oxidized state of the A-cluster, the [Fe₄S₄] cubane, the Ni_p, and the Ni_d atoms are all diamagnetic with charges of +2.^{2,4,9} However, to catalyze acetyl-CoA synthesis, reductive activation of the A-cluster is required to form one of several proposed states: [Fe₄S₄]²⁺–Ni_p⁰–Ni_d²⁺,¹⁰ [Fe₄S₄]¹⁺–Ni_p¹⁺–Ni_d²⁺,¹¹ and [Fe₄S₄]²⁺–Ni_p¹⁺–Ni_d²⁺.^{5,12} Only the latter state is EPR active (*S* = 1/2), and this paramagnetic species (termed the *NiFeC species*) has only been observed when the enzyme is incubated with CO.^{13,14} It has remained an unresolved question, addressed here, whether the active carbonylated form of the A-cluster is the paramagnetic or one of the diamagnetic metal–carbonyl (M–CO) states. The aim of these studies is to measure the rate of formation of the M–CO species by stopped-flow infrared spectroscopy (SF–IR)¹⁵ and to determine whether this rate coincides with the rate of formation of the paramagnetic *NiFeC species* by rapid-freeze quench EPR (RFQ–EPR). Both rates are compared with that of steady-state catalysis to determine catalytic competence.

A monofunctional ACS has been isolated from *Carboxydothermus hydrogenoformans* (ACS_{CH}).⁶ We cloned and over-expressed ACS_{CH} in *E. coli* (see Supporting Information) to yield a purified ACS_{CH} containing 3–4 Fe atoms per monomer but no Ni. Upon reconstitution with NiCl₂ and treatment with CO and DTH, ACS_{CH} exhibits the *NiFeC* EPR signal with typical *g* values of 2.067 and 2.026 (0.38–0.67 spins/ACS_{CH} monomeric unit). The sample used in the IR experiment had 0.67 spins/ACS_{CH} (indicating

that at least 67% of ACS_{CH}–CO is paramagnetic) and exhibited the following activities (at 30 °C): CO/acetyl-CoA exchange, 0.25 U/mg (0.34 s^{–1}); acetyl-CoA synthesis, 0.029 U/mg (0.04 s^{–1}); and dephosphoCoA/acetyl-CoA exchange,¹⁶ 23 U/mg (32 s^{–1}).

To explore CO binding to ACS, SF–IR experiments were performed at 25 °C by rapidly mixing 100 μM ACS_{CH} with CO-saturated buffer, with both solutions containing 2 mM DTH and 2 mM DTT (Figure 1). When ACS_{CH} is reacted with ¹²CO, the only significant metal–CO band that forms at a rate consistent with catalysis is at 1994.7 cm^{–1}. A similar band observed with the *M. thermoacetica* CODH/ACS was assigned to a terminally coordinated Ni–CO.^{17,18} When ACS_{CH} is reacted with ¹³CO, the band shifts 45 cm^{–1} to 1949.7 cm^{–1}, which is consistent with the 44.3 cm^{–1} predicted using a simple harmonic oscillator for a bound CO. The band increases with time of CO incubation, following a single exponential with a rate constant of 0.83 ± 0.06 s^{–1} (Figure 2). A weaker band forms at a rate much slower than that of catalysis at 2044 cm^{–1} (3% of the 1994.7 cm^{–1} band after 800 s). When DTH is absent, the same M–CO bands are seen, but these form much more slowly, presumably due to the weaker reducing power of DTT compared to that of DTH.

When the bifunctional CODH/ACS is reacted with CO, at least five bands are observed with stretching frequencies in the range of 2200–1800 cm^{–1}. One of these bands (at 1996 cm^{–1}) was proposed to correspond to the A-cluster of ACS, and the others were assigned to CODH.^{17,18} Observation of a similar band at 1994.7 cm^{–1} with the monofunctional ACS_{CH} unambiguously demonstrates that the 1996 cm^{–1} band indeed arises from the carbonylated A-cluster and implies that the other bands are correctly assigned to M–CO complexes associated with the C-cluster in CODH/ACS.

To determine whether the M–CO band corresponds to a diamagnetic or paramagnetic state, RFQ–EPR was performed in which 80 μM ACS_{CH} was reacted with CO at 22 °C under otherwise identical conditions as the SF–IR experiments. The *NiFeC* signal developed with a rate constant of 1.4 ± 0.2 s^{–1} (Figure 2). The *NiFeC* species has *g* values identical to those of native *C. hydrogenoformans* ACS⁶ and similar to those of the CODH/ACS from *M. thermoacetica* and methanogens.^{13,19}

That the 1994.7 cm^{–1} IR band and the *NiFeC* EPR signal form at similar rates and are the predominant species present in solution strongly indicates that they represent the vibrational and magnetic spectral fingerprints of the same M–CO species. Furthermore, the IR band and the *NiFeC* signal develop faster than the rates of acetyl-CoA synthesis and the CO/acetyl-CoA exchange reactions, which satisfies the criterion of catalytic competence of the M–CO species in these reactions. When reacted with the methylated CF₂SP, the paramagnetic *NiFeC* species decays 6-fold faster than the steady-state catalytic rate,⁵ further satisfying the criterion of catalytic

[†] Lawrence Berkeley National Laboratory.

[‡] University of Nebraska.

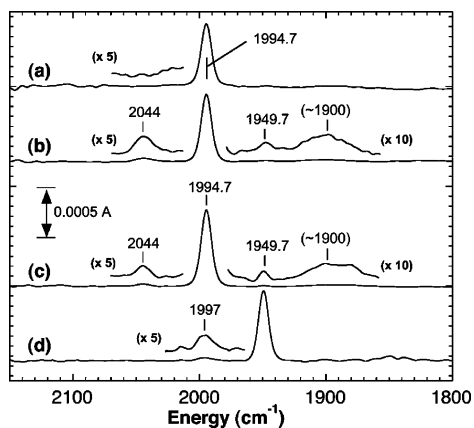


Figure 1. SF-IR spectra of 50 μM ACS_{CH} (final) reacted with CO: (a) 3.0 and (b) 877 s after reaction with ^{12}CO in the presence of 2 mM DTH. Reaction after 1100 s in the absence of DTH (c) ^{12}CO , (d) ^{13}CO . The small band at 1949.7 cm^{-1} in (b) and (c) corresponds to natural abundance ^{13}CO (1.07%). The collection time for (a) was 1 s. The others were collected over 20 s.

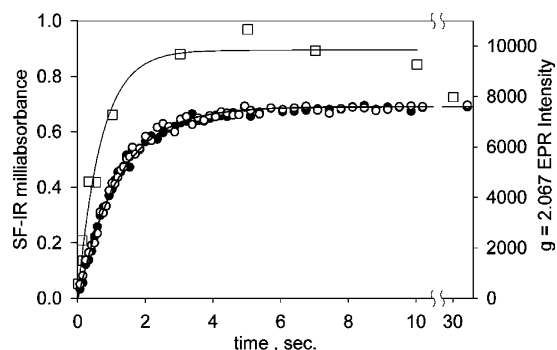


Figure 2. Reaction of ACS_{CH} with CO at 25 °C followed by FQ-EPR and SF-IR. Formation of the 1994.7 cm^{-1} band in two separate experiments is shown as closed and open circles. The data fit to a single exponential with $k_{\text{obs}} = 0.83 \text{ s}^{-1}$ (solid line). The $g = 2.067$ EPR signal (open squares) followed a single exponential with $k_{\text{obs}} = 1.4 \text{ s}^{-1}$. EPR spectra were recorded as described in the Supporting Information.

competence. That the M-CO complex involves $\text{Ni}^{1+}\text{-CO}$ is supported by FT-IR,¹⁷ EPR,¹³ ENDOR,²⁰ and computational studies.²¹

We do not observe reduction of the $[\text{Fe}_4\text{S}_4]^{2+}$ by UV/visible or EPR spectroscopy when DTH alone is added to Ni-reconstituted ACS_{CH} (data not shown). This agrees with previous EPR, EXAFS, and Mössbauer experiments, indicating the NiFeC species contains a diamagnetic $[\text{Fe}_4\text{S}_4]^{2+}$ cluster.²² Thus, a $[\text{Fe}_4\text{S}_4]^{2+}\text{-Ni}^{1+}\text{-CO}$ is supported for the IR band at 1994.7 cm^{-1} . Since the only observed carbonylated form of ACS_{CH} is a *paramagnetic* complex, diamagnetic $\text{Ni}^{2+}\text{-CO}$ and $\text{Ni}^0\text{-CO}$ states are unlikely intermediates in acetyl-CoA synthesis.

Acknowledgment. We gratefully acknowledge NIH (GM39451) for support, and Prof. Roger N.F. Thorneley, John Innes Centre, Norwich, U.K., for access to the SF-FTIR spectrometer.

Supporting Information Available: Description of the cloning and overexpression of ACS_{CH} into pQE70, description of the assays and methodology for SF-IR and RFQ-EPR, as well as original EPR and IR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- (1) Abbreviations: CODH/ACS, CO dehydrogenase/acetyl-CoA synthase; ACS_{CH}, acetyl-CoA synthase from *Carboxydotherrnus hydrogenoformans*; SF-IR, stopped-flow Fourier transformed infrared spectroscopy; RFQ-EPR, rapid-freeze-quench EPR; DTT, dithiothreitol; DTH, dithionite.
- (2) Ragsdale, S. W. *Crit. Rev. Biochem. Mol. Biol.* **2004**, *39*, 165–195.
- (3) (a) Drake, H. L. *Acetogenesis*; Chapman & Hall: New York, 1994; pp 3–60. (b) Ferry, J. G. *Annu. Rev. Microbiol.* **1995**, *49*, 305–333.
- (4) Lindahl, P. A. *Biochemistry* **2002**, *41*, 2097–2105.
- (5) Seravalli, J.; Kumar, M.; Ragsdale, S. W. *Biochemistry* **2002**, *41*, 1807–1819.
- (6) Svetlitchnyi, V.; Dobbek, H.; Meyer-Klaucke, W.; Meins, T.; Thiele, B.; Romer, P.; Huber, R.; Meyer, O. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 446–451.
- (7) (a) Doukov, T. I.; Iverson, T. M.; Seravalli, J.; Ragsdale, S. W.; Drennan, C. L. *Science* **2002**, *298*, 567–572. (b) Darnault, C.; Volbeda, A.; Kim, E. J.; Legrand, P.; Vernede, X.; Lindahl, P. A.; Fontecilla-Camps, J. C. *Nat. Struct. Biol.* **2003**, *10*, 271–279.
- (8) (a) Drennan, C. L.; Doukov, T. I.; Ragsdale, S. W. *J. Biol. Inorg. Chem.* **2004**, *9*, 511–515. (b) Lindahl, P. A. *J. Biol. Inorg. Chem.* **2004**, *9*, 516–524.
- (9) (a) Riordan, C. *J. Biol. Inorg. Chem.* **2004**, *9*, 542–549. (b) Brunold, T. C. *J. Biol. Inorg. Chem.* **2004**, *9*, 533–41.
- (10) (a) Lindahl, P. A. *J. Biol. Inorg. Chem.* **2004**, *9*, 516–524. (b) Webster, C.; Darensbourg, M.; Lindahl, P.; Hall, M.B. *J. Am. Chem. Soc.* **2004**, *126*, 3410–3411.
- (11) Gencic, S.; Grahame, D. A. *J. Biol. Chem.* **2003**, *278*, 6101–6110.
- (12) (a) Gorst, C. M.; Ragsdale, S. W. *J. Biol. Chem.* **1991**, *266*, 20687–20693. (b) Kumar, M.; Lu, W.-P.; Liu, L.; Ragsdale, S. W. *J. Am. Chem. Soc.* **1993**, *115*, 11646–11647.
- (13) Ragsdale, S. W.; Wood, H. G.; Antholine, W. E., *Proc. Natl. Acad. Sci. U.S.A.* **1985**, *82*, 6811–6814.
- (14) Shin, W.; Lindahl, P. A. *Biochemistry* **1992**, *31*, 12870–12875.
- (15) Thorneley, R. N. F.; George, S. J. In *Prokaryotic Nitrogen Fixation: A Model System For Analysis of a Biological Process*; Triplett, E. W., Ed.; Horizon Scientific Press: Wymondham, U.K., 2000.
- (16) This partial reaction measures the rapid rate of CoA binding and C-S bond formation/cleavage, but does not assess catalytic competence of the M-CO species, an intermediate in C-C bond formation.
- (17) Cheng, J.; Huang, S.; Seravalli, J.; Gutzman, H.-J.; Schwartz, D.; Ragsdale, S. W.; Bagley, K. *Biochemistry* **2003**, *42*, 14822–14830.
- (18) Kumar, M.; Ragsdale, S. W. *J. Am. Chem. Soc.* **1992**, *114*, 8713–8715.
- (19) Lu, W.-P.; Jablonski, P. E.; Rasche, M.; Ferry, J. G.; Ragsdale, S. W. *J. Biol. Chem.* **1994**, *269*, 9736–9742.
- (20) Fan, C.; Gorst, C. M.; Ragsdale, S. W.; Hoffman, B. M. *Biochemistry* **1991**, *30*, 431–435.
- (21) Schenker, R. P.; Brunold, T. C. *J. Am. Chem. Soc.* **2003**, *125*, 13962–13963.
- (22) (a) Xia, J.; Hu, Z.; Popescu, C. V.; Lindahl, P. A.; Munck, E. *J. Am. Chem. Soc.* **1997**, *119*, 8301–8312. (b) Russell, W. K.; Stalhandske, C. M. V.; Xia, J.; Scott, R. A.; Lindahl, P. A. *J. Am. Chem. Soc.* **1998**, *120*, 7502–7510.

JA0528329